

# **The Auburn University Detection and Food Safety Center: *Our Mission and Accomplishments***



AUBURN UNIVERSITY

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DETECTION AND FOOD  
SAFETY CENTER



**December 2010**

**Bryan A. Chin, Director**

# **AUDFS is a multi-disciplinary, multi-college initiative.**

**Samuel Ginn  
College of Engineering**

**College of  
Agriculture**



**College of  
Sciences and  
Mathematics**

**AUBURN UNIVERSITY**

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**DETECTION AND FOOD  
SAFETY CENTER**

**College of  
Veterinary Medicine**

**College of Human  
Sciences**

## Research Team (Past and Present)

### College of Agriculture

- Donald E. Conner, Professor and Chair  
HAACP, Sampling
- Jacek Wower, Professor  
RNA/DNA
- Omar Oyarzabal, Associate Professor  
Food Sample Preparation, Food Microbiology
- Manpreet Singh, Assistant Professor  
Food Science, Microbiology
- Tung-Shi Huang, Assistant Professor  
Antibody Development, Immobilization
- Jean Weese, Assoc. Professor/Ext. Food Specialist  
Industrial Practices, Needs

### Samuel Ginn College of Engineering

- ZhongYang Cheng, Associate Professor  
Polymers, Magnetostrictive Devices
- Bryan A. Chin, Professor and Chair  
Bulk Food Monitor, Bacteria/Spore Binding
- Barton C. Prorok, Associate Professor  
MEMs Design, M<sup>3</sup>S Devices
- Dong-Joo (Daniel) Kim, Assistant Professor  
MEMs Fabrication, Microcantilevers
- Aleksandr L. Simonian, Associate Professor  
SPR, Bio-chemical recognition
- Jeffrey W. Fergus, Associate Professor  
Carbon Dioxide Bacterial Sensing
- Jong Wook Hong, Assistant Professor  
Microfluidic Devices
- Jin Wang, Assistant Professor  
Statistical Probability
- William F. Gale, Professor  
Education, Metal Surfaces/Bacteria Interaction
- Jeff Smith, Professor  
Food/Plant Security
- Yonhua Tzeng, Professor  
Microelectronics, Thin films
- Mark Byrne, Associate Professor  
Kinetics, Polymer Membranes, Drug Delivery

### College of Human Sciences

- Hae-jung An, Assistant Professor  
Food Science, Sampling
- Peggy Hsieh, Professor  
Test Kits, Food Species Determination
- Cheng-I Wei, Professor  
Food Science, Food Microbiology

### College of Sciences and Mathematics

- William Charles Neely, Professor  
Air/Liquid Extraction, Chemical Binding
- Curtis Shannon, Assoc. Professor  
Self Molecular Assembly
- James M. Barbaree, Professor  
Microbiology, Bacteria, Spores
- Minseo Park, Assistant Professor  
Raman Spectroscopy
- Sang-Jin Suh, Assistant Professor  
Phage Development
- Wei Zhan, Assistant Professor  
Biosensors, Chemical Detection
- Laura Suh, Assistant Professor  
Spores, Food Safety
- Robert Locy, Professor  
Plant Biochemistry, Signal Transduction
- An-Ban Chen, Professor  
Modeling
- David Held, Assistant Professor  
Food/Plant Security

### College of Veterinary Medicine

- Vitaly Vodyanoy, Professor  
Molecular Recognition, Immobilization
- Valery A. Petrenko, Professor  
Bio-molecular Recognition, Phage
- Arnold Vainrub, Associate Professor  
Bio-molecular Recognition, DNA

Auburn University

Detection and  
Food Safety  
Center

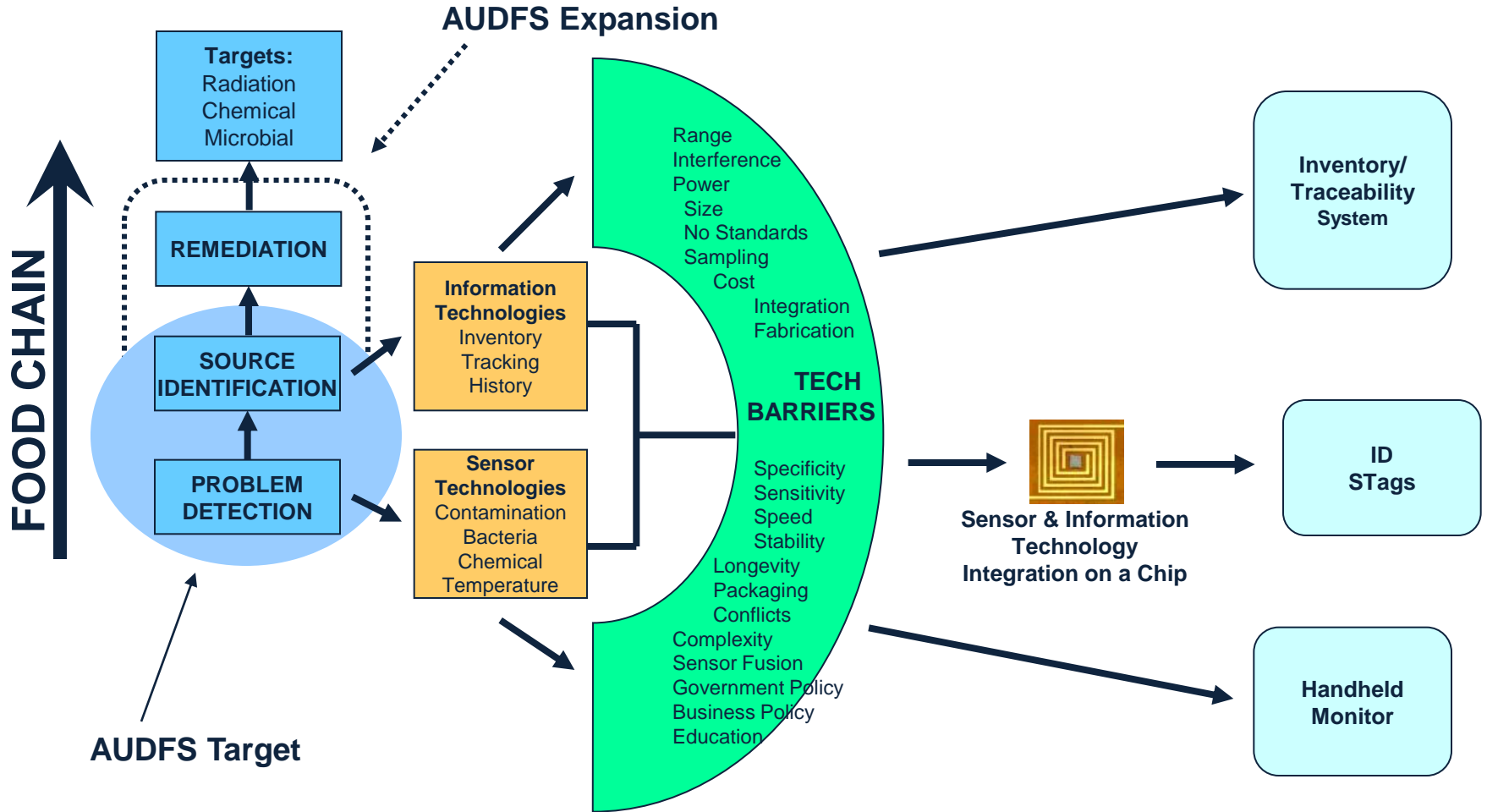
## Auburn University Detection and Food Safety Center

Our vision is to improve the safety of the U.S. food system by developing the science and engineering required to rapidly identify, pinpoint and characterize problems that arise in the food supply chain through the integration of sensor and information systems technology.

[www.auburn.edu/audfs](http://www.auburn.edu/audfs)



# AUDFS Focus



# AUDFS Annual Assessment

	2004	2005	2006	2007	2008	2009	2010
<b>Publications (Refereed Pubs) (cumulative)</b>	132	152	181	239	285	324	354
<b>Students, Staff, Research Assoc. supported (annual)</b>	55	56	112	54	66	59	54
<b>M.S. Graduates (cumulative)</b>	28	38	44	46	48	57	61
<b>Ph.D. Graduates (cumulative)</b>	23	34	39	42	54	67	74
<b>Patents</b>	9	10	15	19	19	20	
<b>Disclosures</b>	36	54	89	109	113	119	133
<b>Commercialized Products</b>	3	4	4	5	6	6	6

# Commercialized Technologies

	2004	2005	2006	2007	2008	2009	2010
<b>Disclosures (cumulative)</b>	36	54	89	109	113	119	133
<b>Patents Awarded (cumulative)</b>	9	10	15	19	19	20	
<b>Commercialized Technology (cumulative)</b>	3	4	4	5	6	6	6



**Raytheon RFID Bulk STag for Patriot Missile Health Monitoring Mil Spec Qualified**



**Meat species identification technology licensed to ELISA Technologies**



**RMBM in livestock feed identification technology licensed to Neogen Corp.**



**Optical microscope technology licensed to Aetos Technologies, Inc.**



**Canine Olfactory Measurement Method licensed to RedXDefense, Inc.**

## Requirements of AUDFS Participants

- Two refereed journal publications per year
- One AUDFS PhD graduate every 5 years
- Use AUDFS funding to gather preliminary data for grant application
- One externally funded grant every three years that exceeds AUDFS cumulative funding by 4 times

# Spinoff Center: FAA Center of Excellence for Airliner Cabin Environment Research (ACER)



- 8th institution center with AU as Administrative Lead
- \$3.5 M from FAA in first year of operation
- 1:1 matching by industry & member universities
- 26 industry partners & growing
- Major focus on chem.–bio. sensing aboard airliners
  - Cross-membership between AUDFS and ACER teams
  - ACER is implementing technologies developed by AUDFS
  - ACER is bringing new industry partners to AUDFS
- ACER is now part of the new National Center of Excellence for Research in the Intermodal Transport Environment (RITE)

# Spinoff Project: Condition Based Maintenance for Military Vehicles



**Contract: \$1 million/year**

**Sponsored by FTI Inc. through the U.S. Army TACOM CBM**

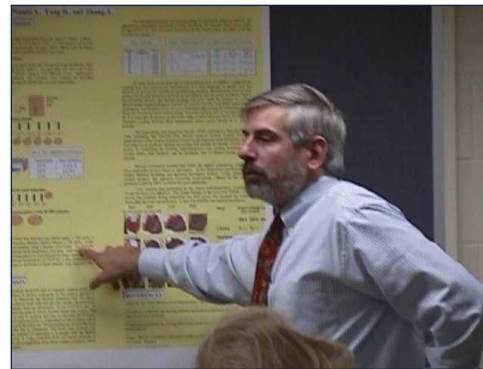
- Project will produce a universal vehicle condition monitoring system based on AUDFS-developed technologies (shipment and plant monitoring system) that enables predictive and proactive maintenance
- Employs sensor concepts and data acquisition/recording hardware developed by AUDFS to monitor critical parameters related to vehicle condition and reliability
- Project seeks to reduce maintenance costs and provide early warning of impending failures of expensive battle equipment, affording soldiers better protection from terrorists and enemy fighters
- Works toward implementation of a microscale device to monitor engine oil conditions in a “smart dipstick”

# Cross-disciplinary Food Science-Engineering Course

- Over \$1 Million in Funded Education Projects (USDA & NSF)
- Streaming Video and DVD for Off-Campus Students
- Food Safety/Production Practitioners
- Credit for Certification and Advanced Degrees



Profs. Bill Gale &  
Jeff Fergus



Profs. Don Conner &  
Omar Oyarzabal



# AUDFS TECHNOLOGIES

# Ongoing AUDFS Investigations

- RNA-based biosensors for pathogen detection
- New biodetection method using amplified photo electrochemical signaling
- Air sampling for *E. coli* detection on spinach leaves
- Chemical sensing strategies based on molecularly imprinted polymers
- Wireless amperometric sensors for pathogen detection
- FET DNA biosensors for pathogen detection
- Improvement of phage probes using genetic engineering
- Phage for MRSA detection
- Immobilizing phage using Langmuir-Blodgett Films
- Surface modification of ME materials to allow direct immobilization of phage
- Liquid phage solutions for cleaning of food preparation surfaces
- Vortex Centrifuges for detection of contamination in large volumes of food
- Invasive insect species detection using Smart automated traps
- Magnetoelastic sensors for pathogen detection on fresh fruits/vegetables
- Magnetoelastic sensors for in-situ monitoring of plant physiology

# Phage-based Magnetoelastic Biosensor

Landscape Phage as Bio-receptor

Magnetoelastic Particle as Sensor Platform

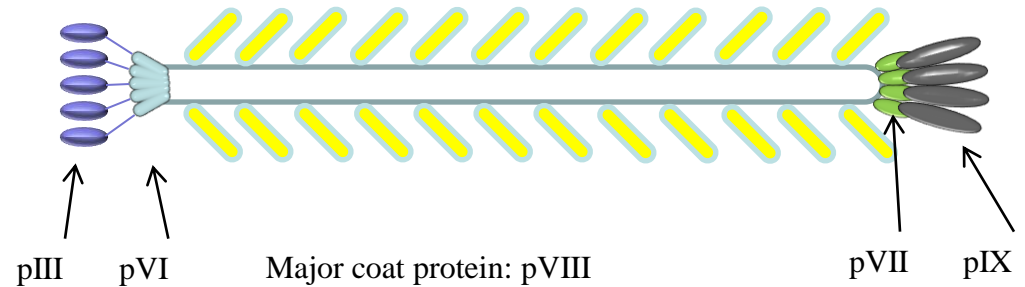
**JRB7 Phage** against *Bacillus anthracis*

**E2 Phage** against *Salmonella typhimurium*

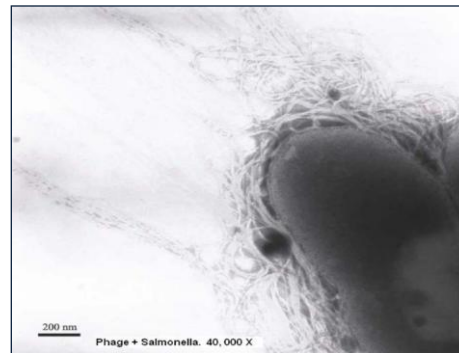
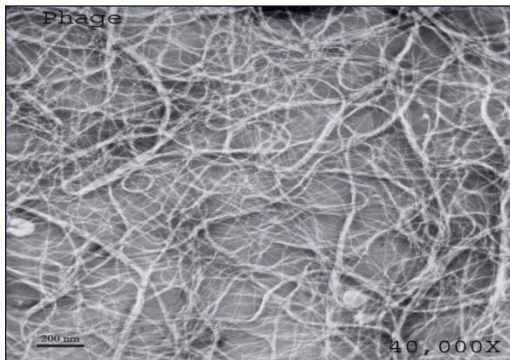
# Biomolecular Recognition

## Filamentous Phage

- Virus-like agents that infect bacteria
- Investigating filamentous phages including M13, f1 and f3
- Approximately 7nm diameter and 900 nm long
- Genetically engineered coat proteins for specificity



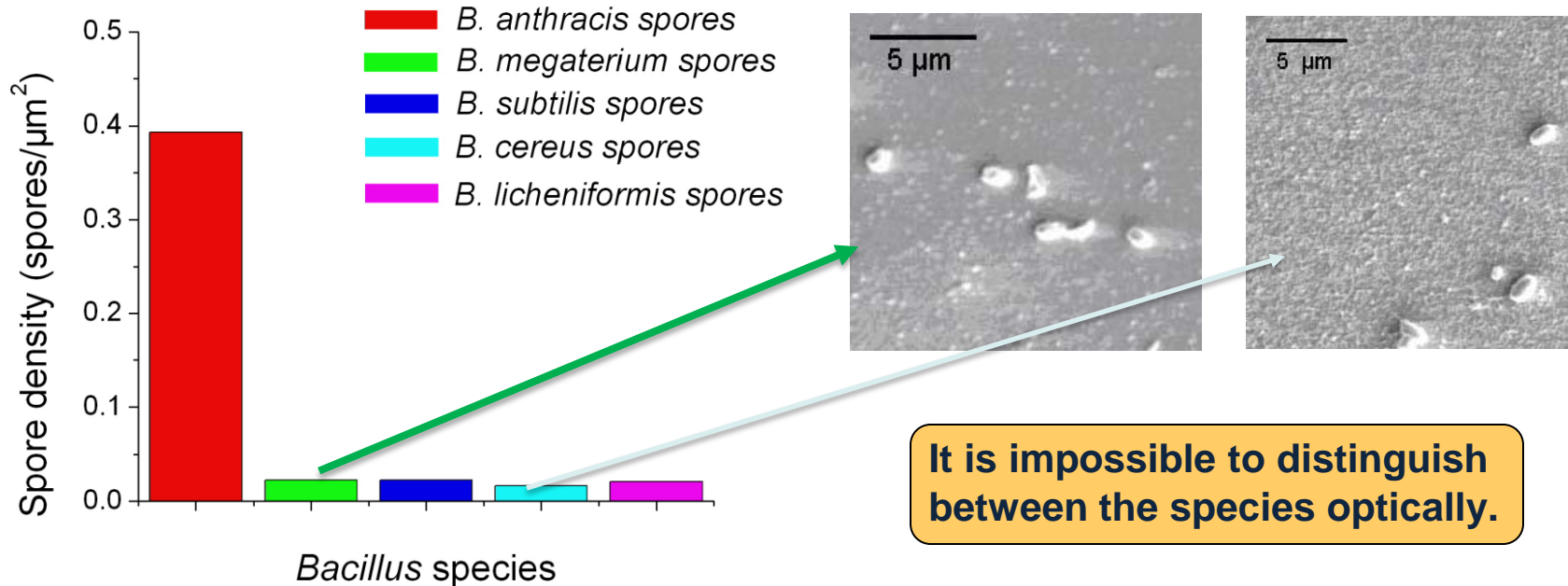
## TEM micrographs



## AUDFS Patents

- *Bacillus anthracis* phage
- *Salmonella* phage

## Specificity of *B. Anthracis* Phage

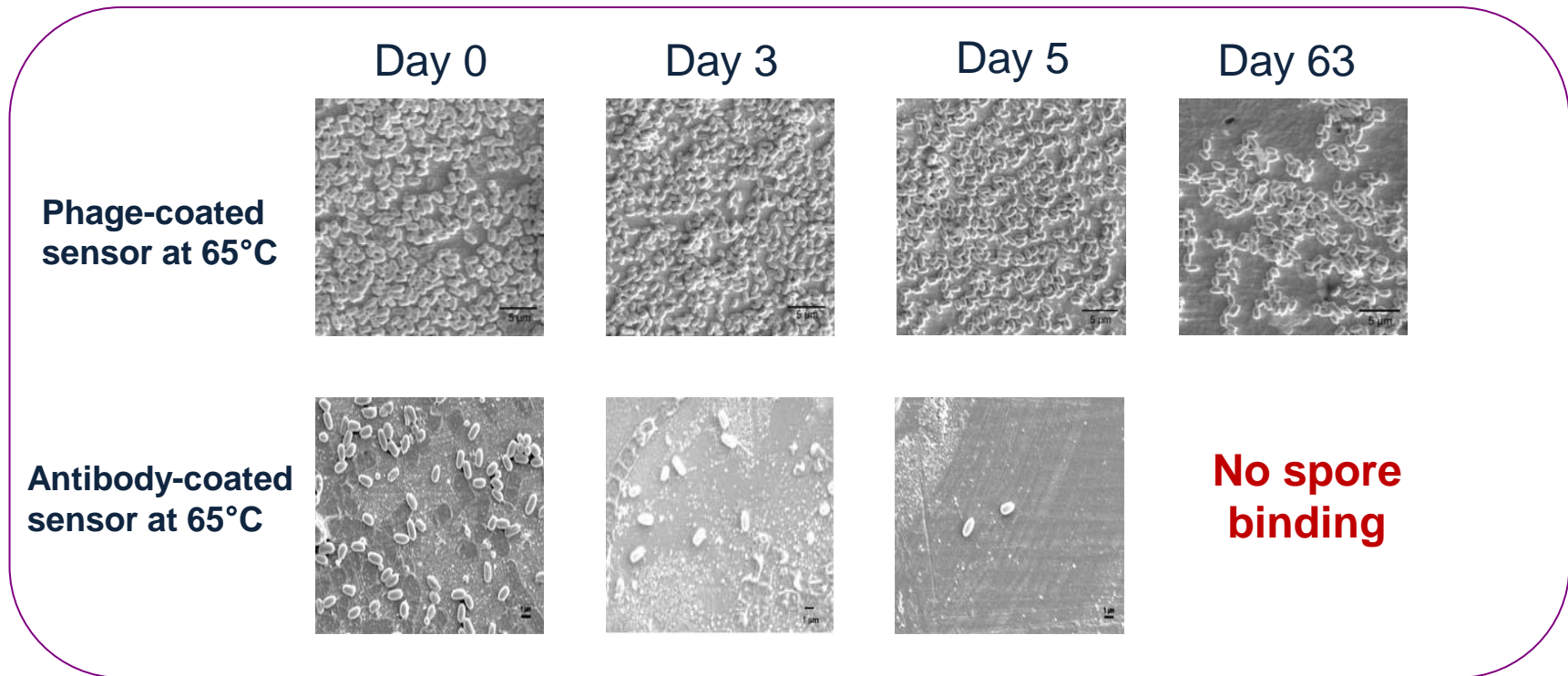


***Bacillus Anthracis* Spores bound phage-coated magnetoelastic sensors**

- 40-fold better than *B. licheniformis* and *B. megaterium*
- 15-fold better than *B. subtilis* and *B. cereus* spores.

# Longevity of Phage-coated Sensors

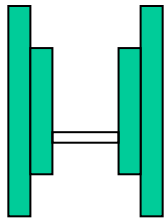
The SEM images of phage-coated sensors and antibody-coated sensors at 65°C



- The binding activity of antibody-coated biosensor dropped to zero after storage at 65°C for 5 days.
- Phage-coated biosensors still showed good binding affinity after two months.

# Magnetoelastic Particle Sensor

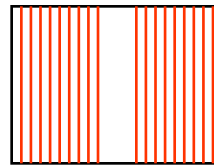
**Driving Coil**



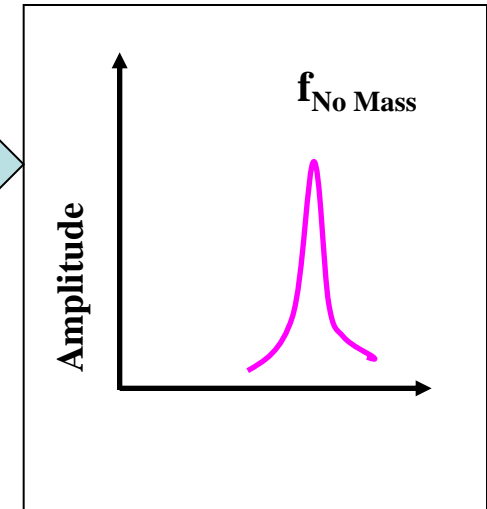
**Applied  
Varying  
Magnetic  
Field**



**Pick-Up Coil**



**Resultant  
Magnetic  
Field Signal**



**Magnetoelastic  
Resonator**

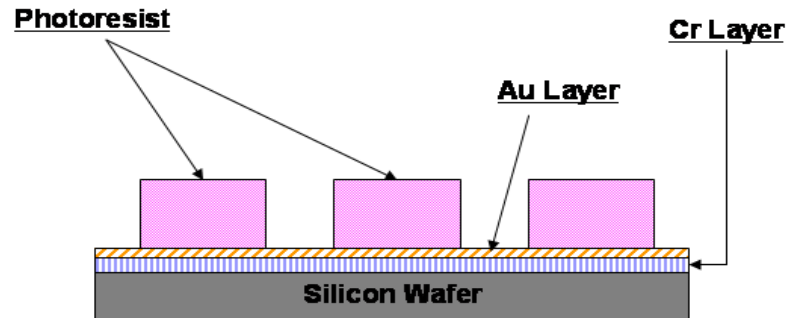
**Phage/Antibody or  
Chemical Binding layer**

**Longitudinal  
Oscillation**

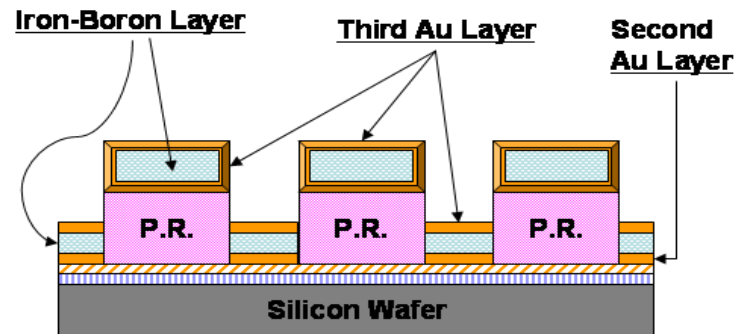


# Fabrication of Magnetoelastic Sensors

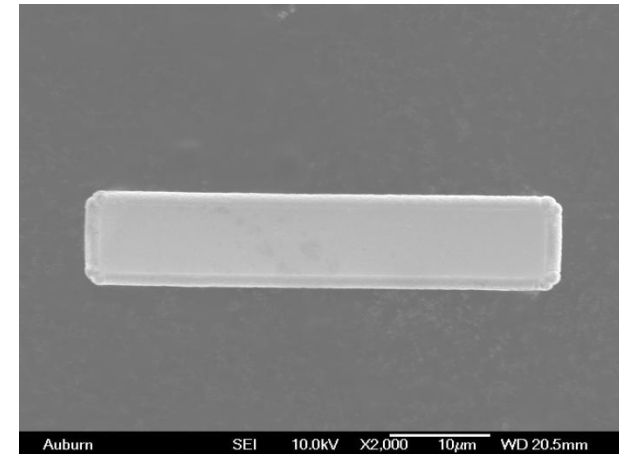
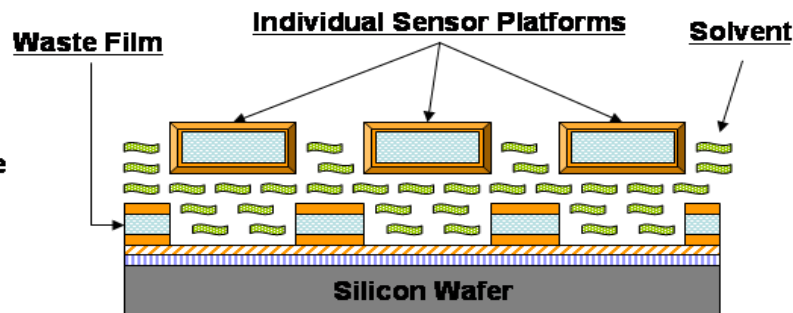
1. Patterning of the chromium and gold coated silicon wafer.



2. Deposition of a second gold layer, the magnetoelastic iron-boron layer, and finally a top gold layer.



3. Lift-off of the particles is performed by dissolving the photoresist with a solvent, leaving the surrounding waste film on the wafer.



4 x 10 x 50 µm Sensor  
Capable of Detecting  
a Few Spores

## Size of the ME Biosensor

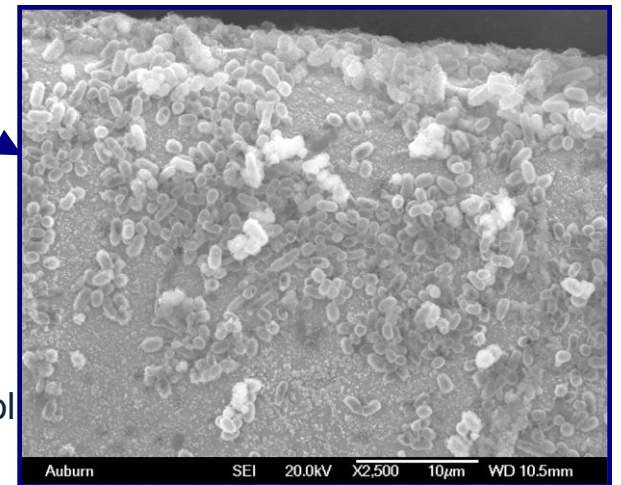
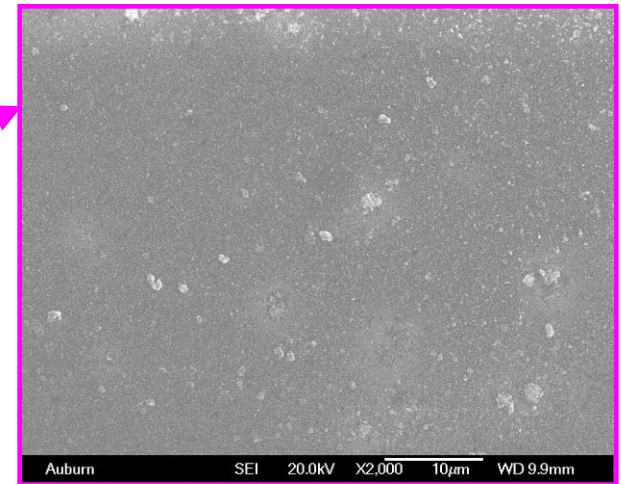
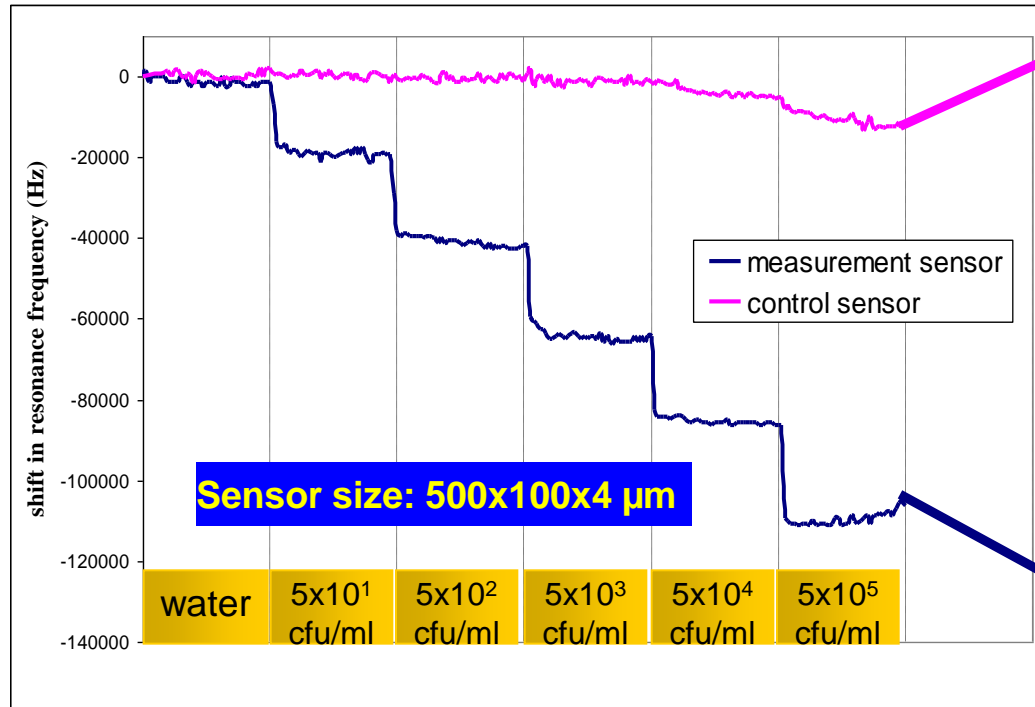


Scanning Electron Micrograph comparing the size of a ME biosensor with the Y in "LIBERTY" on a penny.

The biosensors are microelectronically fabricated and are smaller than a particle of dust.

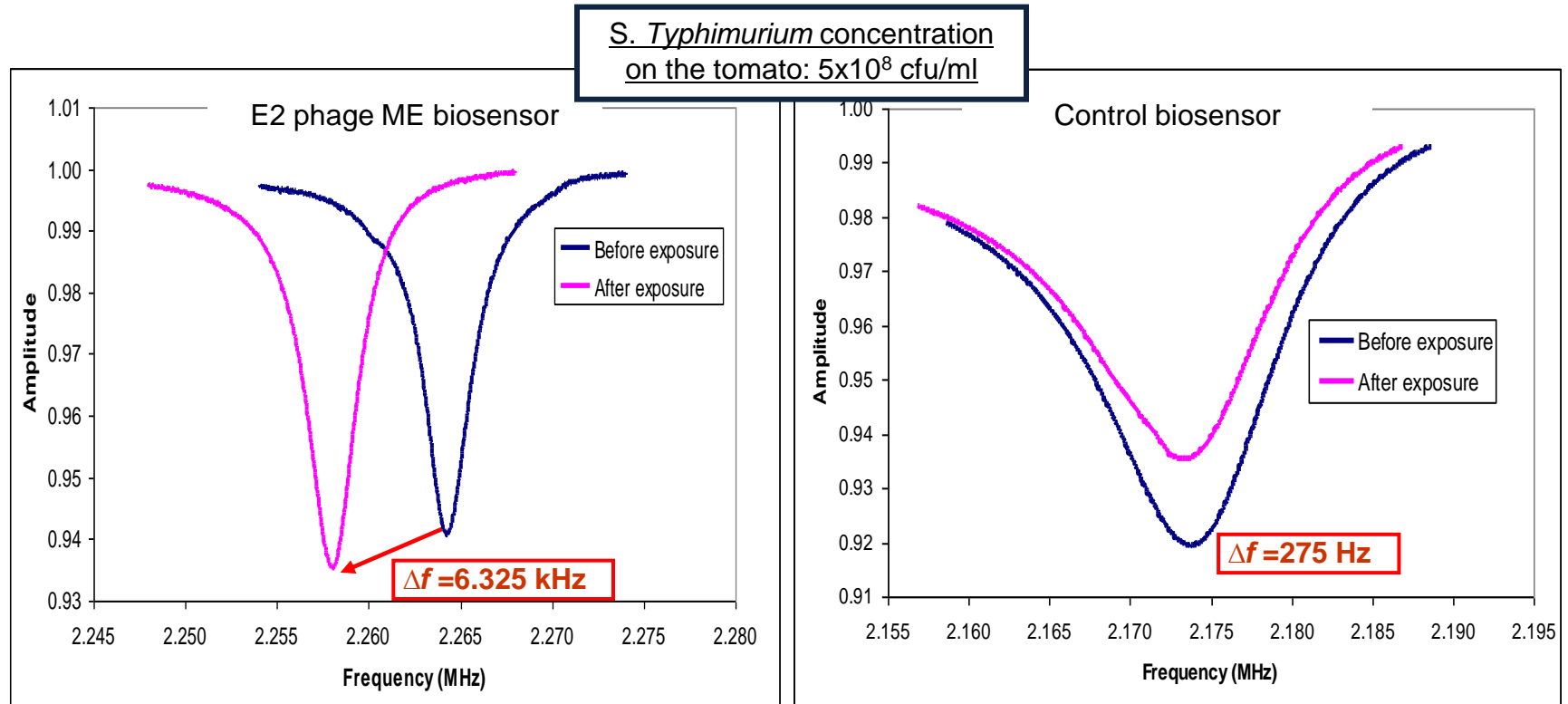
The biosensors require no on-board power and their cost is less than 1/1000 of a cent each when fabricated in large numbers.

## Response of 500 micron long biosensor exposed to increasingly higher concentrations of *Salmonella*



Detection limit better than 50 CFU/ml. The response of the control sensor (devoid of phage) is also shown. The SEM images show near zero binding of *Salmonella* cells to the control sensor and a large number of bound *Salmonella* cells to the measurement sensor.

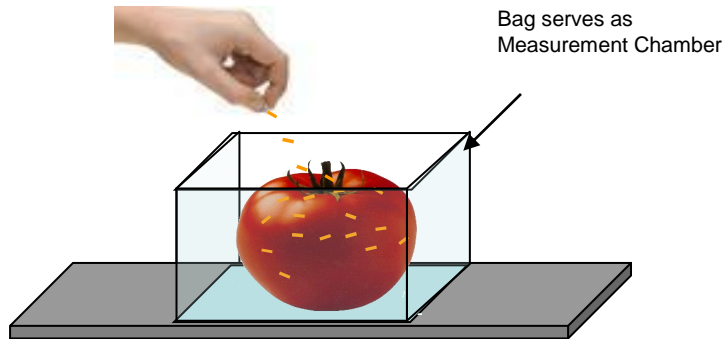
# *S. typhimurium* Detection on Tomato Surface



Resonance Frequencies of Control and Measurement Biosensors  
After Exposure to Tomato Spiked with *S. typhimurium* ( $5 \times 10^8$  cfu/mL)

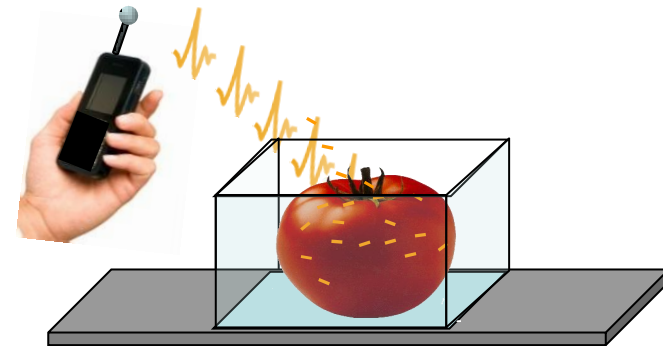
# Vision of Direct Bacterial Detection on Foods

## 1 Distribute Sensors (1 minute)



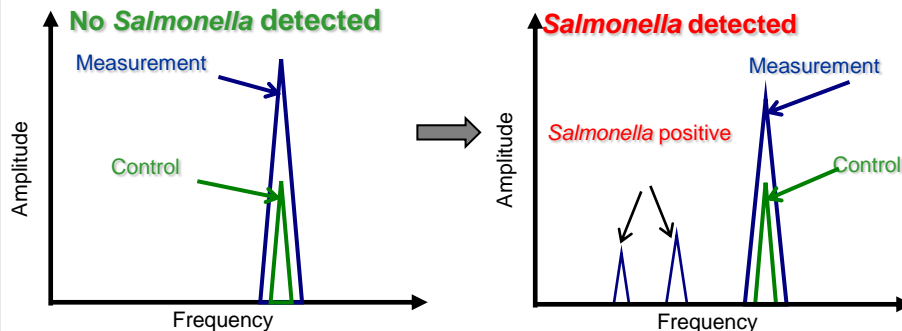
Sprinkle 55 sensors (50 measurement, 5 control) onto the tomato, wait 1 minute for binding to occur.

## 2 Measure (< 1 minute)



Measure wirelessly and simultaneously the 55 sensors. Less than 1 minute.

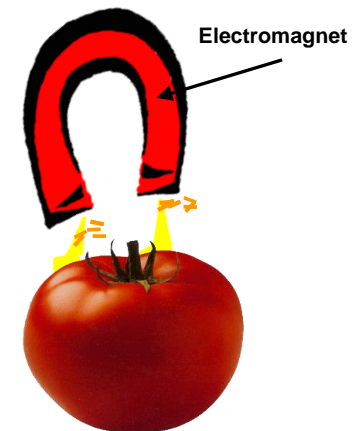
## 3 Analyze Data (< 1 minute)



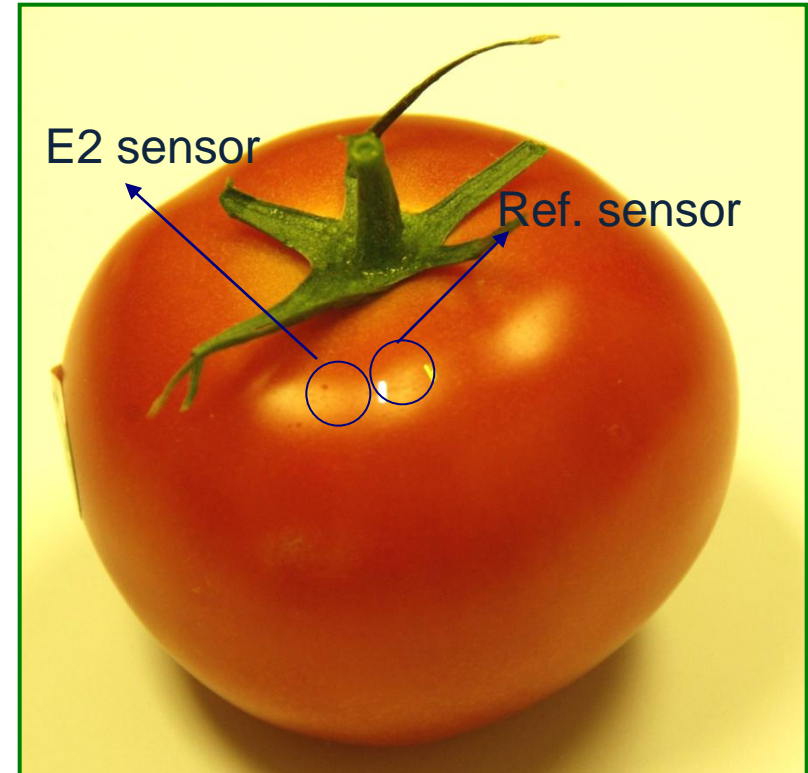
If control and measurement sensors show same resonance frequency, then an undetectable amount of *Salmonella* is present.

If peaks appear below control, then have positive *Salmonella* contamination.

## 4 Retrieve Sensors (< 1 minute)

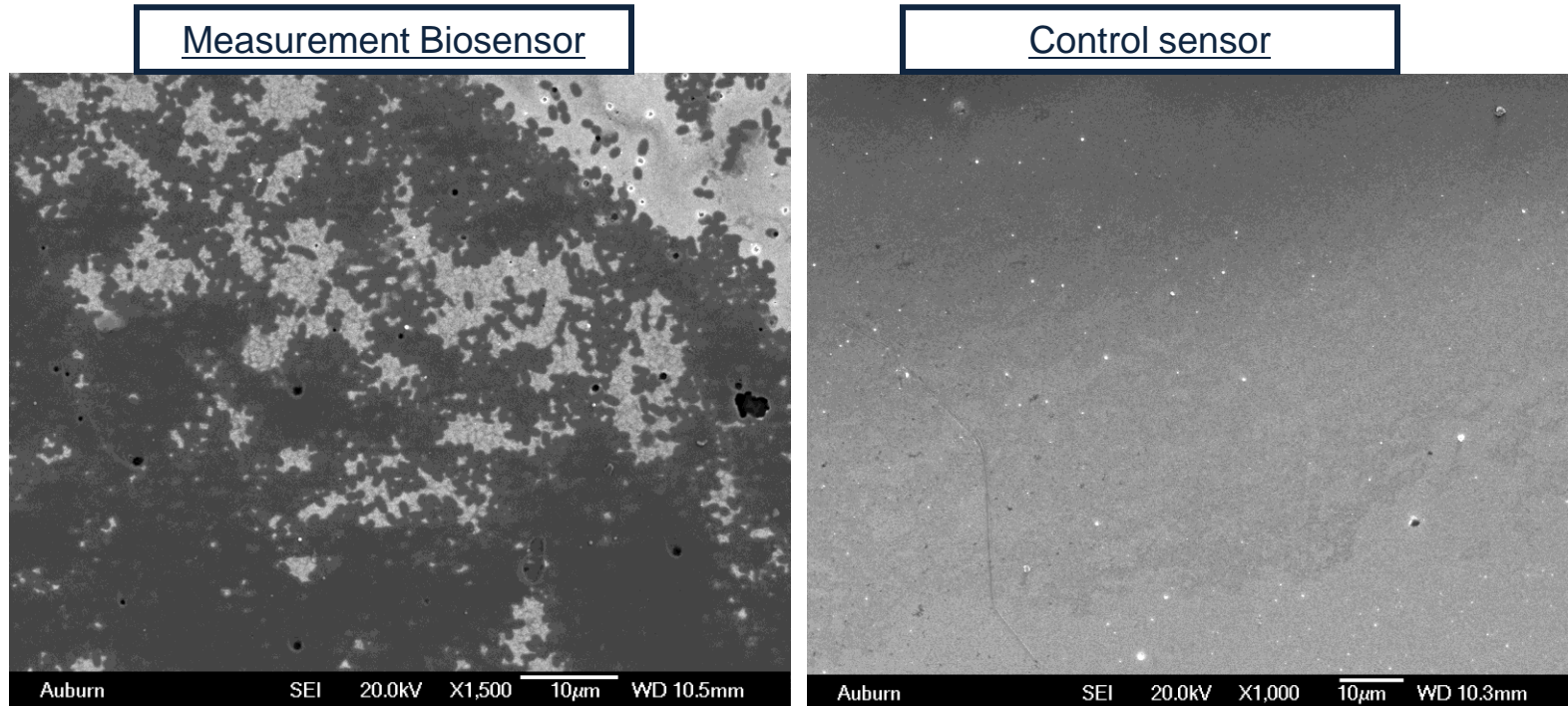


## ***S. typhimurium* Detection on Tomato Surface**



Fresh tomato → Put 20 $\mu$ l *S. typhimurium* suspension and let it dry in air → Place E2 phage-coated ME biosensor on the dried tomato for 30 mins at 85% humidity

## ***S. typhimurium* Detection on Tomato Surface**



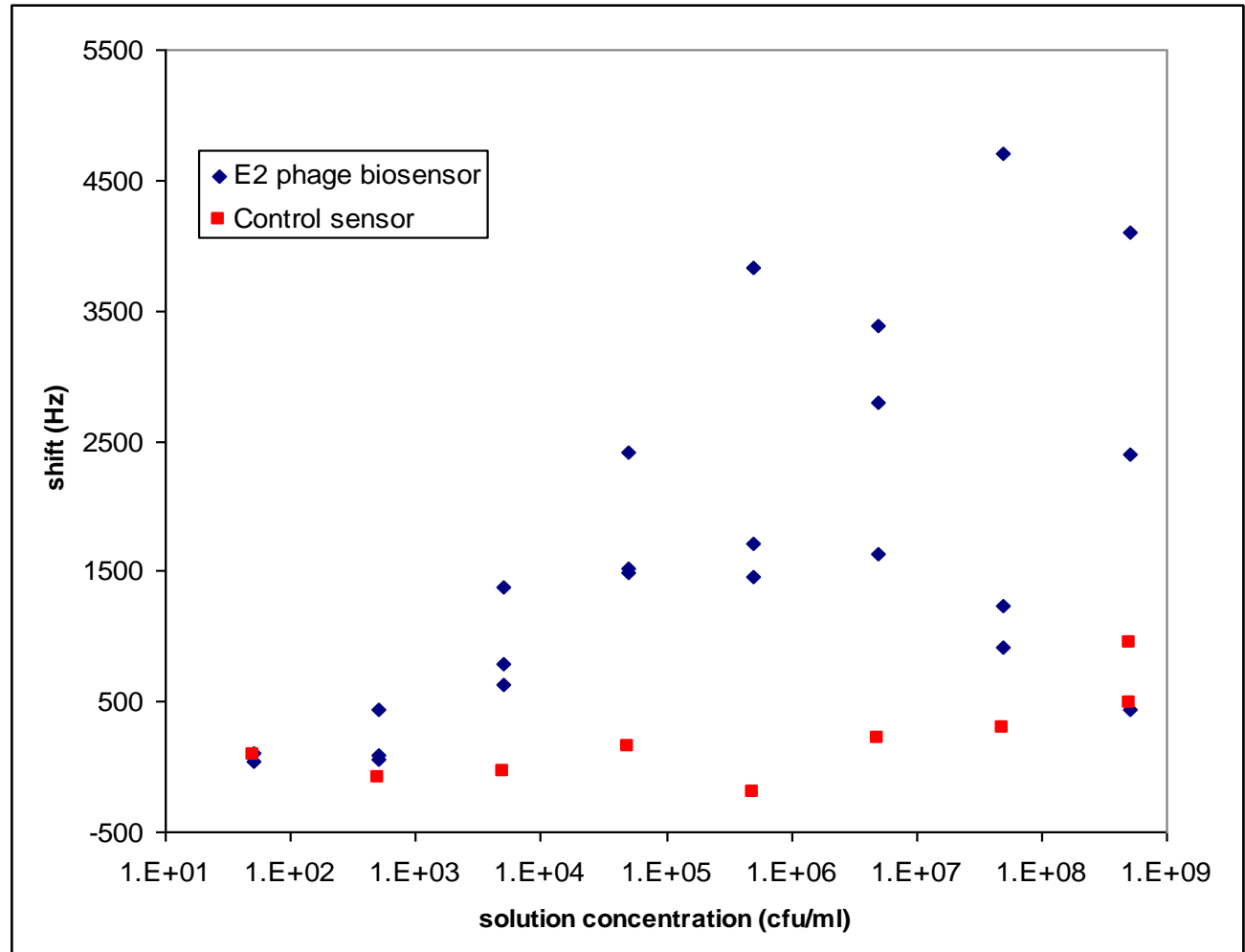
Surface of Control and Measurement Bioensors

After Exposure to Tomato Spiked with *S. typhimurium* ( $5 \times 10^8$  cfu/mL)

# Statistical Response of *S. typhimurium* Detection on Tomato Surface

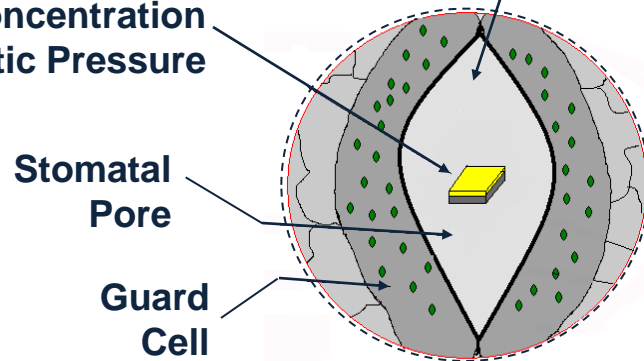
Sensor size:  
1000 x 200 x 28  $\mu\text{m}$

Each data point is  
the average of  
three sensors.

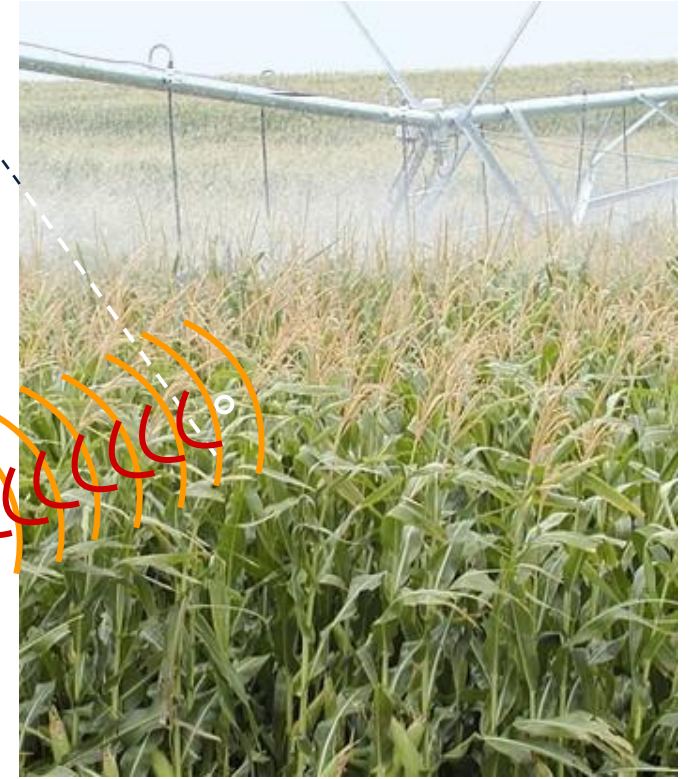
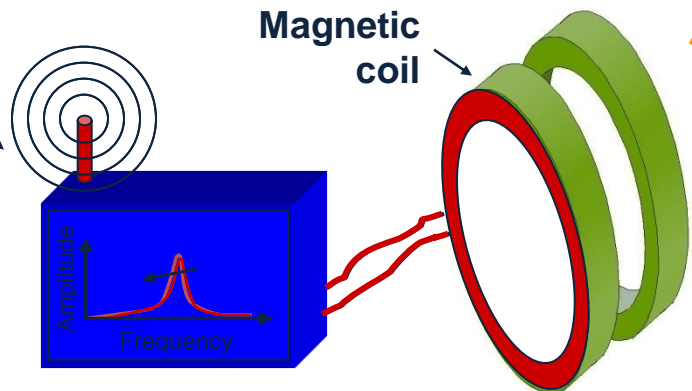


# Internal Plant Growth Monitor

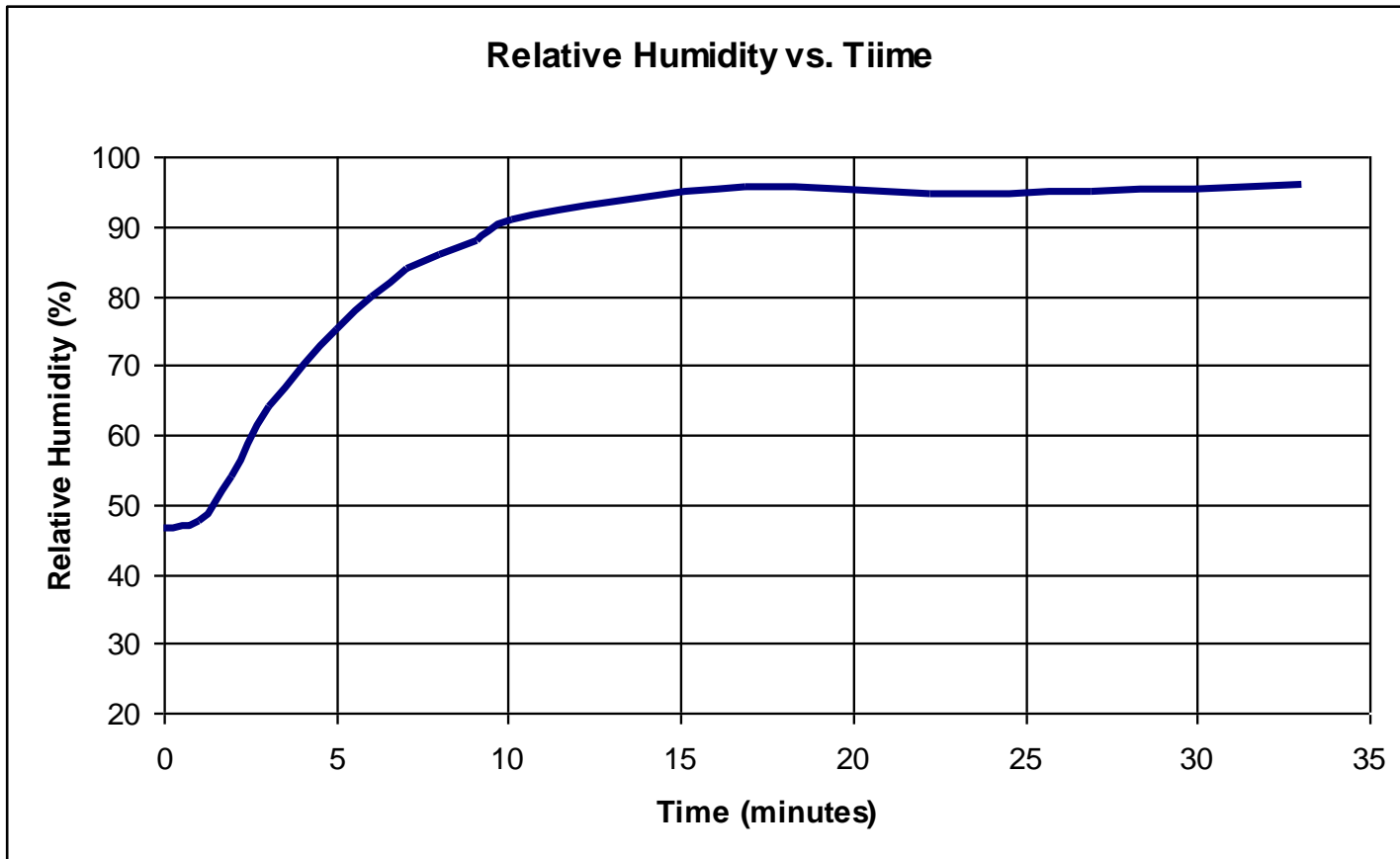
- Magnetoelastic Sensor**
- Temperature
  - Humidity
  - Ion Concentration
  - Osmotic Pressure



**Processing Electronics and Transceiver**



# Internal Cavity Humidity as Measured by Magnetoelastic Sensor



# Questions?



Funded by:

United States  
Department of  
Agriculture

National Institute  
of Food and  
Agriculture